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# Studies on chromium(III) complexes with active nitrogen, oxygen and sulfur donor ketimines synthesized under microwave conditions

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Heterocyclic ketimines, 1-(2-furanyl)ethanonehydrazincarbothioamide (L<sup>1</sup>H), 1-(2-furanyl)ethanonehydrazincarboxamide (L<sup>2</sup>H), 1-(2-furanyl)ethanone hydrazincarbothioamide (L<sup>3</sup>H) and 1-(2-thienyl) ethanonehydrazincarboxamide (L<sup>4</sup>H), were prepared by the condensation of thiosemicarbazide and semicarbazide hydrochloride (in the presence of sodium acetate) in ethanol with the respective ketones by using microwave as well as conventional methods. Chromium(III) complexes have been prepared by mixing CrCl<sub>3</sub> · 6H<sub>2</sub>O in 1:1 and 1:2 molar ratios with monobasic bidentate ketimines. The authenticity of the ligands and their complexes has been established by elemental analyses, melting point determinations, molecular weight determinations, EPR, infrared and UV spectral and X-ray powder diffraction studies. These studies showed that the ligands coordinated to the metal atom in a monobasic bidentate mode, coordinating through the nitrogen and sulfur/oxygen donor system. Thus, an octahedral environment around the metal atoms has been proposed. The growth-inhibiting potential of the ligands and complexes has been assessed against a variety of fungal and bacterial strains.

Keywords: chromium(III) complexes; heterocyclic ketimines; spectral studies; antimicrobial activity

### 1. Introduction

Increasing attention for environmental protection during the last decades has led both modern academic and industrial groups to develop chemical processes with the maximum yield and minimum cost while using non-toxic reagents, solvents and catalysts (1). Microwave (MW) synthesis represents one of the important dimensions of modern chemistry attracting a considerable amount of attention (2). Metal complexes, in particular of transition metal ions, are important in many areas of science, including catalysis, medicine (diagnosis and therapy), design of high value materials, analytical chemistry and as model compounds with the structure and function of metalloproteins (3, 4). The metal oxidation state, the type and number of donor atoms, as well as their relative disposition within the ligand, are major factors determining the structure–activity relationship of the metal complexes (5). Chromium(III) is a unique transition metal, which has been established

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to be biologically significant at all the levels of living organisms (6). Out of the two stable oxidation states of chromium, chromium(III) has been shown to play a positive role in controlling carbohydrate and lipid metabolism (7). For instance, even at the prokaryotic level, chromic chloride has been shown to maintain the glucose levels in the yeast, *Saccharomyces cerevisiae* (8). Ketimine complexes of chromium(III) are efficient inhibitors for the pathogenic microorganism and have invasive potential of *Shigella dysenteriae* (9). Thiosemicarbazones usually act as chelating ligands with the transition metal ion, bonding through the sulfur and azomethine nitrogen atoms. The chemistry of thiosemicarbazones has received considerable attention in view of their variable bonding modes, promising biological implications, structural diversity and ion-sensing ability (10). Semicarbazones and thiosemicarbazones have been used as drugs and are reported to possess a wide variety of biological activities against bacteria, fungi and certain types of tumors, and they are also a useful model for bioinorganic processes (11).

Because of the wide range of medicinal applications of semicarbazones and thiosemicarbazones and their abilities to coordinate with the transition metal ions, it is highly desirable to synthesize and characterize the transition metal complexes with such ketimines. Keeping all these facts under consideration during the present investigations, we have synthesized, characterized and screened the biologically potent ligands and their chromium(III) complexes against a variety of pathogenic fungal and bacterial strains.

## 2. Results and discussion

The resulting chromium(III) complexes are green solids, soluble in MeOH, DMF and DMSO and sparingly soluble in  $H_2O$ . Molecular weight determinations indicate their monomeric nature. The reactions of  $CrCl_3 \cdot 6H_2O$  with the ligands were carried out in unimolar and bimolar ratios in methanol. The successive replacement of chloride resulted in the formation of products  $[CrCl_2(L)(H_2O)_2]$  and  $[(CrCl(L)_2(H_2O)]$  as shown in Equations (1) and (2).

$$CrCl_3 \cdot 6H_2O + LH + NaOH \xrightarrow[MeOH]{1:1} [CrCl_2(L)(H_2O)_2] + NaCl + 5H_2O$$
(1)

$$CrCl_3 \cdot 6H_2O + 2LH + 2NaOH \xrightarrow{1:2}_{MeOH} [(CrCl(L)_2(H_2O)] + 2NaCl + 7H_2O.$$
(2)

The physical properties and analytical data of the ligands and their metal complexes, synthesized by green chemical approach as well as conventional method, are enlisted in Tables 1 and 2.

#### 2.1. Electronic spectra

The electronic spectra of the complexes were recorded in DMSO. In the electronic spectra of chromium(III) complexes, three transitions are expected and are also observed experimentally. A consistent treatment for the bands is to assign the band near 15,550–17,900 cm<sup>-1</sup> as the  ${}^{4}A_{2}g \rightarrow {}^{4}T_{2}g(\nu_{1})$  transition; the band at 22,850–24,040 cm<sup>-1</sup> as the  ${}^{4}A_{2}g \rightarrow {}^{4}T_{1}g(\nu_{2})$  transition and that at 32,700–30,080 cm<sup>-1</sup> as the  ${}^{4}A_{2}g \rightarrow {}^{4}T_{1}g(P)(\nu_{3})$  transition. These three transitions suggest an octahedral geometry around the Cr<sup>+3</sup> ion (*12*, *13*). Various ligand field parameters such as Dq, *B*, and  $\beta$  have been calculated and given in Table 3. The energy of the first spin-allowed transition [ ${}^{4}A_{2}g(F) \rightarrow {}^{4}T_{2}g(F)$ ] directly gives the value of 10Dq. The ligand field spectral parameters for chromium(III) complexes are found to be Dq = 1790–1680 cm<sup>-1</sup>, *B* = 471–882 cm<sup>-1</sup> and  $\beta$  = 0.51–0.96. The reduction in the Racah parameter (*B*) from the free-ion value of 918–654 cm<sup>-1</sup> and the value of  $\beta$  indicate the covalent nature of the chromium(III) complexes.

		Molting		Found	d (calculate	d) (%)		Molecular weight found
Compounds	Color	point (°C)	С	Н	Ν	S	М	(calculated)
L <sup>1</sup> H	Brown	108-110	45.01	4.87	22.09	16.99	_	186.30
			(45.88)	(4.91)	(22.93)	(17.50)		(183.23)
$L^{2}H$	White	160-162	49.95	4.96	25.71	_	_	169.75
			(50.29)	(5.42)	(25.14)			(167.09)
L <sup>3</sup> H	Yellow	130-132	41.85	4.21	20.90	32.96	_	205.37
			(42.18)	(4.55)	(21.08)	(32.17)		(199.30)
$L^{4}H$	White	187-190	45.01	4.42	22.15	17.18	-	185.32
			(45.88)	(4.95)	(22.93)	(17.50)		(183.23)
$[CrCl_2(L^1)(H_2O)_2]$	Green	140-143	24.14	3.67	11.84	10.97	15.44	343.32
			(24.62)	(3.54)	(12.03)	(9.39)	(15.29)	(341.14)
$[CrCl(L^1)_2(H_2O)]$	Green	160-164	34.77	3.92	17.98	13.06	10.91	471.31
			(35.78)	(3.86)	(17.03)	(13.64)	(11.07)	(469.89)
$[CrCl_2(L^2)(H_2O)_2]$	Green	262-265 (d)	25.96	3.51	10.80	_	16.10	329.25
			(25.84)	(3.72)	(11.08)		(16.18)	(325.07)
$[CrCl(L^2)_2(H_2O)]$	Green	223–225 (d)	38.78	3.75	15.78	_	10.13	496.36
			(38.11)	(4.14)	(15.30)		(10.97)	(491.26)
$[CrCl_2(L^3)(H_2O)_2]$	Green	180–200 (d)	22.66	2.84	12.03	17.52	14.86	360.46
			(23.52)	(3.38)	(12.11)	(17.97)	(14.56)	(357.01)
$[CrCl(L^3)_2(H_2O)]$	Green	215-225 (d)	32.45	3.10	16.07	25.17	10.72	505.34
			(33.49)	(3.61)	(16.74)	(25.54)	(10.37)	(501.03)
$[CrCl_2(L^4)(H_2O)_2]$	Green	190–200 (d)	23.43	3.88	12.25	9.86	15.00	346.34
			(24.64)	(3.52)	(12.69)	(9.39)	(15.29)	(341.14)
$[CrCl(L^4)_2(H_2O)]$	Green	150–160 (d)	36.21	3.13	18.09	13.11	11.79	471.19
			(35.84)	(3.86)	(17.88)	(13.65)	(11.07)	(469.90)

Table 1. Analytical data and physical properties of the ligands and their complexes synthesized by conventional heating.

Note: d, decomposition.

Table 2.	Analytical da	ta and physical	properties of	f the ligands and	their complexes	synthesized by N	AW method.
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		Melting		Found (calculated) (%)					
Compounds	Color	point (°C)	С	Н	Ν	S	М	(calculated)	
L <sup>1</sup> H	Brown	108-110	45.12	4.97	22.87	17.46	_	184.51	
			(45.88)	(4.91)	(22.93)	(17.50)		(183.23)	
$L^{2}H$	White	160-162	50.15	5.89	25.10	-	-	168.99	
			(50.29)	(5.42)	(25.14)			(167.05)	
L <sup>3</sup> H	Yellow	130-132	41.83	4.52	21.45	32.12	-	200.55	
			(42.18)	(4.55)	(21.08)	(32.17)		(199.30)	
$L^{4}H$	White	187-190	46.01	4.88	22.91	17.38	-	185.90	
			(45.88)	(4.95)	(22.93)	(17.50)		(183.23)	
$[CrCl_2(L^1)(H_2O)_2]$	Green	140-143	24.54	3.60	12.56	9.31	15.78	342.22	
			(24.62)	(3.54)	(12.03)	(9.39)	(15.29)	(341.14)	
$[CrCl(L^1)_2(H_2O)]$	Green	160-164	34.77	3.05	17.00	13.46	11.15	470.76	
			(35.78)	(3.86)	(17.03)	(13.64)	(11.07)	(469.89)	
$[CrCl_2(L^2)(H_2O)_2]$	Green	262–265 (d)	25.92	3.01	11.24	-	16.09	327.42	
			(25.84)	(3.72)	(11.08)		(16.18)	(325.07)	
$[CrCl(L^2)_2(H_2O)]$	Green	223–225 (d)	37.74	3.97	15.16	_	10.81	492.25	
			(38.11)	(4.14)	(15.30)		(10.97)	(491.26)	
$[CrCl_2(L^3)(H_2O)_2]$	Green	180–200 (d)	23.66	3.05	12.19	17.83	14.72	359.92	
			(23.52)	(3.38)	(12.11)	(17.97)	(14.56)	(357.01)	
$[CrCl(L^3)_2(H_2O)]$	Green	215-225 (d)	32.99	3.58	16.57	25.68	10.21	503.64	
			(33.49)	(3.61)	(16.74)	(25.54)	(10.37)	(501.03)	
$[CrCl_2(L^4)(H_2O)_2]$	Green	190–200 (d)	24.43	3.49	12.69	9.46	15.33	343.83	
			(24.64)	(3.52)	(12.69)	(9.39)	(15.29)	(341.14)	
$[CrCl(L^4)_2(H_2O)]$	Green	150–160 (d)	35.11	3.37	17.29	13.69	10.95	470.88	
		. ,	(35.84)	(3.86)	(17.88)	(13.65)	(11.07)	(469.90)	

Note: d, decomposition.

Commenceda	Tar	Spectral bands	De	DO	0 0 00	
Compounds	Transitions	cm <sup>-</sup> (nm)	Dq	В	$p = B/B^{*}$	$v_2/v_1$
$[CrCl_2(L^1)(H_2O)_2]$	$\label{eq:A2g} \begin{array}{l} {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (P) \end{array}$	16,800 (595) 23,100 (432) 30,080 (332)	1680	617	0.67	1.37
$[CrCl(L^1)_2(H_2O)]$	$\label{eq:A2g} \begin{array}{l} {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (P) \end{array}$	17,340 (576) 23,950 (417) 32,050 (312)	1734	651	0.70	1.38
$[CrCl_2(L^2)(H_2O)_2]$	$\label{eq:A2g} \begin{array}{l} {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g}\left(F\right) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}\left(F\right) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}\left(P\right) \end{array}$	15,550 (643) 22,995 (434) 30,314 (329)	1555	882	0.96	1.47
$[CrCl(L^2)_2(H_2O)]$	$\label{eq:A2g} \begin{array}{l} {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (P) \end{array}$	17,540 (570) 24,040 (415) 32,356 (309)	1754	635	0.69	1.37
$[CrCl_2(L^3)(H_2O)_2]$	${}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g}(F)$ ${}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}(F)$ ${}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}(P)$	17,900 (553) 23,050 (433) 31,500 (317)	1790	471	0.51	1.28
$[CrCl(L^3)_2(H_2O)]$	${}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g}(F)$ ${}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}(F)$ ${}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}(P)$	17,210 (581) 23,000 (434) 30,300 (330)	1721	552	0.60	1.33
$[CrCl_2(L^4)(H_2O)_2]$	$\label{eq:4} \begin{array}{l} {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g}\left(F\right) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}\left(F\right) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g}\left(P\right) \end{array}$	17,380 (575) 23,290 (429) 32,700 (306)	1739	563	0.61	1.34
$[CrCl(L^4)_2(H_2O)]$	$\label{eq:A2g} \begin{array}{l} {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{2g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (F) \\ {}^{4}A_{2g}(F) \rightarrow {}^{4}T_{1g} \ (P) \end{array}$	17,035 (586) 22,850 (436) 31,780 (315)	1704	563	0.61	1.34

Table 3. Electronic spectral data  $(cm^{-1})$  of the chromium(III) complexes.

Note: B, Complex; Bo, free ion.

#### 2.2. ESR spectra and magnetic moment

The electron spin resonance spectra of 1:1 and 1:2 chromium(III) complexes were recorded at room temperature. These consist of a single broad peak in each case and from which the Lande splitting factor ("g" values) has been calculated which lie in the range 1.9289–1.9431, with  $g_{iso}$  (2.04) which are characteristic of an octahedral geometry (14). The room temperature magnetic moment for the chromium(III) chelates is slightly less than the required environment. The observed magnetic moment value of 3.92–4.20 BM and the electronic spectra of the complexes support the octahedral structure of the complexes (15).

### 2.3. Infrared spectra

The infrared (IR) spectra of the thiosemicarbazones (L<sup>1</sup>H and L<sup>3</sup>H) and semicarbazones (L<sup>2</sup>H and L<sup>4</sup>H) display a sharp band at 1610–1600 cm<sup>-1</sup>, which is shifted to a lower frequency, in the spectra of the corresponding metal complexes, probably due to the coordination of the azomethine nitrogen to the metal atom. In the spectra of the free ligands, bands appearing due to  $\nu$ (NH) vibrations in the regions 3240–3230 cm<sup>-1</sup> disappear in the spectra of the complexes, indicating deprotonation of the functional groups on complexation. Medium-intensity bands due to  $\nu$ (C=S)/ $\nu$ (C=O) vibrations appear in the regions 1050–1040/1690–1680 cm<sup>-1</sup> in the spectra of ligands and shifted to the lower frequency in the spectra of the complexes. This fact is further corroborated by the observation of the new bands due to  $\nu$ (C–O) and  $\nu$ (C–S) modes at lower frequency in the spectra of chromium(III)

complexes. In the spectra of chromium(III) complexes, a band is observed in the range 870–860 cm<sup>-1</sup>, which may be attributed to the coordinated water molecule. Further, a broad band around 3480–3400 cm<sup>-1</sup>may be due to  $\nu$ (O–H) of water molecule. The far IR spectra of these metal complexes exhibited new bands, which are not present in the spectra of the ligands. The single band observed at ca. 320–315 cm<sup>-1</sup> is due to  $\nu$ (Cr–Cl), suggesting thereby that the complexes have a *trans* structure (*16*) and the presence of  $\nu$ (Cr–N),  $\nu$ (Cr–S) (*17*) and  $\nu$ (Cr–O) vibrations around 535 ± 10, 360–335 and 610 ± 10 cm<sup>-1</sup> indicate that complexation takes place through the nitrogen and sulfur/oxygen atoms.

## **2.4.** NMR spectra $({}^{1}H \text{ and } {}^{13}C)$

The proposed structures of the ligands get further support by the <sup>1</sup>H NMR and <sup>13</sup>C spectra of the ligands. The significant <sup>1</sup>H NMR and <sup>13</sup>C NMR spectral data of the ligands,  $(L^{1}H, L^{2}H, L^{3}H)$  and  $(L^{4}H)$  along with their tentative assignments are reported in Table 4.

On the basis of above studies, octahedral environment around the metal atoms has been proposed, and the expected structures are as shown in Figures 1 and 2.

	<sup>1</sup> H NMR	spectra (δ	, ppm)		<sup>13</sup> C NMR spectra ( $\delta$ , ppm)				
Ligands	$-C \stackrel{CH_3}{=} N -$ proton	NH proton	NH <sub>2</sub> proton	Aromatic protons	Azomethine carbon	Amido/thiolo carbon	Methyl carbon	Aromatic carbon	
$L^1H$	1.8	9.9	2.83	7.80–6.60	178.51	198.71	12.3	150.81, 134.75,	
$L^2H$	1.78	10.68	2.55	7.89–6.32	160.07	177.35	12.33	153.02, 144.19,	
$L^{3}H$	1.68	10.68	2.81	8.68–7.16	146.2	179.78	13.74	143.40, 128.54,	
L <sup>4</sup> H	2.3	10.68	2.84	7.84–7.20	152.76	178.62	13.17	140.31, 126.45, 126.70, 125.37	

Table 4. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectral data ( $\delta$ , ppm) of the ligands.



Figure 1. Suggested structures for chromium(III) complexes. 1:1 complex.



Figure 2. Suggested structures for chromium(III) complexes. 1:2 complex.

## 2.5. X-ray powder diffraction study

The possible lattice dynamics of the finely powdered product,  $[CrCl_2(L^1)(H_2O)_2]$ , has been deduced on the basis of X-ray powder diffraction studies. The observed interplanar spacing values ("d" in Å) have been measured from the diffractogram of the compound (Figure 3); the Miller indices *h*, *k* and *l* have been assigned to each *d* value and  $2\theta$  angles are reported in Table 5. The results show that the compound belongs to "orthorhombic" crystal system having unit cell parameters as a = 9.63, b = 17.37 c = 21.15, maximum deviation of  $2\theta = 0.045^{\circ}$  and  $\alpha = 90^{\circ}$ ,  $\beta = 90^{\circ}$ ,  $\gamma = 90^{\circ}$ .



Figure 3. X-ray powder diffraction spectra of  $[CrCl_2(L^1)(H_2O)_2]$ .

Table 5	5.	X-ray d	iffraction	data	of [C	rCl <sub>2</sub> (L	$L^{1})(H_{2})$	$_{2}O)_{2}].$	

h	k	l	2θ (°) (Exp.)	2θ (°) (Calc.)	2θ (°) (Diff.)	d (Å) (Exp.)	<i>d</i> (Å) (Calc.)	Intensity (Exp.)
0	0	5	26.451	26.476	-0.025	4.23388	4.23000	13.07
0	5	0	32.335	32.380	-0.045	3.47874	3.47400	-3.94
1	0	6	34.071	34.030	0.041	3.30634	3.31020	6.16
2	4	2	36.597	36.650	-0.008	3.08518	3.08453	8.59
0	6	1	39.450	39.475	-0.025	2.86999	2.86825	35.84
1	2	7	41.464	41.468	-0.004	2.73629	2.73607	8.29
1	5	5	43.953	43.995	-0.042	2.58840	2.58604	28.41
1	6	4	46.456	46.469	-0.013	2.45603	2.45537	7.68
4	1	4	52.903	52.922	-0.019	2.17456	2.17384	2.91
0	0	10	54.543	54.514	0.028	2.11358	2.11500	4.88
0	4	9	55.907	55.896	0.011	2.06641	2.06677	11.49
4	4	3	57.379	57.386	-0.007	2.01772	2.01750	28.30
3	7	1	59.402	59.407	-0.005	1.95499	1.95482	9.48
0	9	1	60.571	60.526	0.045	1.92072	1.92201	4.79
4	6	0	63.135	63.106	0.028	1.85032	1.85106	5.82
1	10	0	69.016	69.034	-0.018	1.70980	1.70941	6.13
3	4	10	72.604	72.610	-0.006	1.63610	1.63598	11.05
6	0	1	74.475	74.494	-0.019	1.60074	1.60040	7.37
3	9	4	75.695	75.701	-0.006	1.57861	1.57861	14.29
6	1	4	78.595	78.568	0.027	1.52984	1.52984	7.01
2	3	13	81.164	81.134	0.030	1.48945	1.48945	5.17
4	8	6	82.679	82.694	-0.014	1.46627	1.46627	5.74
3	1	13	84.116	84.105	0.011	1.44614	1.44614	4.87
0	6	13	86.162	86.150	0.012	1.41830	1.41830	7.99

#### 2.6. Antimicrobial assay

The heterocyclic ketimines and their chromium(III) complexes were evaluated for their antimicrobial activity against two bacteria *Bacillus subtilis* (Gram-positive) and *Escherichia coli* (Gram-negative) and two fungi *Candida albicans* and *Aspergillus niger* and the results are summarized in Table 6. The ligands were found to be biologically active and the chromium complexes are more active than their respective ligands. This indicated that the complexation to metal enhances the activity of the ligand. This may be explained by Tweedy's chelation theory (*18*), according to which chelation reduces the polarity of the central metal atom because of partial sharing of its positive charge with the ligand, due to which the lipophilic character of the metal chelate increases and favors its permeation through the lipid layer of cell membrane. It has also been proposed that the ultimate action of the compounds is the denaturation of one or more proteins of the cell as a result of which normal cellular processes are impaired (*19*).

#### 3. Conclusions

MW irradiation is an efficient and environmentally benign method to accomplish various inorganic syntheses to afford purer products in higher yields in shorter reaction periods. All the ligands behave as a monofunctional bidentate with metal ion in different reaction conditions. Based on the analytical and spectral data, a hexacoordinated environment around the metal ion has been proposed. The antimicrobial activity of the complexes and the ligands showed that the complexes are more active than the parent ligands.

### 4. Experimental

The  $CrCl_3 \cdot 6H_2O$  was purchased from Alfa aesar. All the reagents were dried and distilled before use. 2-Acetylfuran and 2-acetylthiophene were purchased and used as received. All preparations were done under anhydrous conditions.

	(Antifungal activity) (%) Inhibition after 96 h (concentration in ppm)					(Antibacterial activity) Diameter (mm) of inhibition zone after 24 h (concentration in ppm)				
	(	C. albica	ns		A. niger		B. sı	ıbtilis	1	E. coli
Compounds	50	100	200	50	100	200	500	1000	500	1000
L <sup>1</sup> H	28	47	50	25	32	52	11	13	10	11
L <sup>2</sup> H	27	44	52	24	30	53	10	11	8	9
L <sup>3</sup> H	28	46	53	26	33	53	11	14	10	13
L <sup>4</sup> H	25	42	51	23	30	50	11	12	9	11
$[CrCl_2(L^1)(H_2O)_2]$	30	49	55	28	35	56	13	14	11	14
$[CrCl(L^1)_2(H_2O)]$	32	50	59	29	38	59	14	15	12	15
$CrCl_2(L^2)(H_2O)_2$ ]	29	49	54	28	36	56	12	13	11	13
$[CrCl(L^2)_2(H_2O)]$	32	52	60	30	42	65	14	13	12	14
$[CrCl_2(L^3)(H_2O)_2]$	30	51	59	31	37	58	12	15	12	16
$[CrCl(L^3)_2(H_2O)]$	33	53	62	32	44	66	15	16	13	16
$CrCl_2(L^4)(H_2O)_2$ ]	29	48	57	27	35	57	13	15	11	13
$[CrCl(L^4)_2(H_2O)]$	31	51	59	30	40	60	14	16	12	15
Nystatin (standard fungicide)	50	85	100	52	89	99	-	-	-	-
Streptomycin (standard bactericide)	-	-	-	-	-	-	16	18	15	18

Table 6. Antifungal and antibacterial screening data for the ligands and their complexes.

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#### 4.1. Preparation of the ligands

Two different routes were employed for the synthesis of the ligands.

- (1) In MW-assisted synthesis, the ligands, thiosemicarbazones (L<sup>1</sup>H, L<sup>3</sup>H) and semicarbazones (L<sup>2</sup>H, L<sup>4</sup>H), were prepared by the condensation of 2-acetylfuran and 2-acetylthiophene with thiosemicarbazide and semicarbazide hydrochloride (in the presence of sodium acetate), respectively, in 1:1 molar ratio through a conventional MW oven by taking 2–4 ml of ethanol as a solvent. The reactions were completed in a short period of 4–7 min.
- (2) For comparison purpose, the above ligands were also synthesized by a thermal method as described previously (20). The parent ligands exist in the tautomeric forms depicted in Figure 4. A comparison between thermal method and MW method is given in Table 7.



#### 4.2. Preparation of the complexes

The complexes were also prepared by two different routes.

(1) In MW-assisted synthesis, the complexes were prepared by irradiating the reaction mixture of chromium trichloride and the respective ligand in appropriate stoichiometric proportions using NaOH in dry methanol. The products were recovered from the MW oven and dissolved in a 2–5 ml of dry methanol, where the precipitate of sodium chloride formed during the course of the reaction was removed by filtration and the filtrate was then concentrated under



Figure 4. Structure of the ligands.

Table 7. Comparison between MW and thermal method.

	Yield (	(%)	Solvent	(ml)	Time		
Compounds	Thermal	MW	Thermal	MW	Thermal (h)	MW (m)	
L <sup>1</sup> H	80	87	100	4	4	5	
L <sup>2</sup> H	79	89	100	3	4	4	
L <sup>3</sup> H	80	86	100	3	4.5	7	
L <sup>4</sup> H	80	88	100	2	3.5	5	
$[CrCl_2(L^1)(H_2O)_2]$	76	83	50	5	15	4	
$[CrCl(L^1)_2(H_2O)]$	73	87	35	2	15	7	
$[CrCl_2(L^2)(H_2O)_2]$	70	89	30	3	14	5	
$[CrCl(L^2)_2(H_2O)]$	72	86	40	2	13	5	
$[CrCl_{2}(L^{3})(H_{2}O)_{2}]$	69	87	55	2	12	4	
$[CrCl(L^3)_2(H_2O)]$	75	81	30	5	15	7	
$[CrCl_2(L^4)(H_2O)_2]$	70	84	30	2	14	7	
$[CrCl(L^4)_2(H_2O)]$	74	80	40	3	15	4	

reduced pressure. The resulting compounds were washed with cyclohexane and recrystallized with methanol.

(2) These complexes were also synthesized by the thermal method where instead of 4–7 min, reactions were completed in 12–15 h and the yield of the products was also less than that obtained by the MW-assisted synthesis. In this method, the methanolic solution of CrCl<sub>3</sub>·6H<sub>2</sub>O was added to the methanolic solution of ligands in 1:1 and 1:2 molar ratios using NaOH in appropriate stoichiometric proportions. The resulting mixture was heated under reflux for 12–15 h, filtered to remove NaCl and the solvent was concentrated under reduced pressure. The product was dried in *vacuum*. The resulting compounds were washed with cyclohexane and recrystallized with methanol.

#### 4.3. Physical measurements and analytical methods

The molecular weights were determined by the Rast Camphor method (21). The metal contents were analyzed gravimetrically. Sulfur and nitrogen were determined by Messenger's (22) and Kjeldahl's methods (23), respectively. Carbon and hydrogen analyses were performed at the CDRI, Lucknow. IR spectra were recorded on a Nicolet Megna FTIR-550 spectrophotometer using KBr pellets. The electronic spectra were recorded on a Varian–Cary/5E spectrophotometer at SAIF, IIT Madras, Chennai. EPR spectra of the complexes were monitored on Varian E-4X band spectrometer at SAIF, IIT Madras, Chennai.

#### 4.4. Antifungal screening

The antifungal activity of the ligands and their chromium complexes was tested by agar plate technique (24) against the two pathogenic fungi *C. albicans* and *A. niger* using the potato dextrose agar medium having the composition: glucose 20 g, starch 20 g, agar–agar 20 g and distilled water 1000 ml. Solutions of the test compounds in methanol at 50, 100 and 200 ppm concentrations were prepared and then were mixed with the medium. The medium then was poured into Petri plates and the spores of fungi were placed on the medium with the help of inoculum's needle. These Petri plates were wrapped in the polythene bags containing a few drops of alcohol and were placed in an incubator at  $25 \pm 2$  °C. The activity was determined after 96 h of incubation at room temperature (25 °C). The controls were also run and three replicates were used in each case. The linear growth of the fungus was obtained by measuring the fungal colony diameter in Petri plates after 4 days. The percentage inhibition was calculated as

% Inhibition = 
$$\frac{(C-T)100}{C}$$
,

where C and T are the diameters of the fungus colony in the control and test plates, respectively.

#### 4.5. Antibacterial screening

*In vitro* antibacterial screening is generally performed by disc diffusion method (25) for the primary selection of the compounds as therapeutic agents. The method is essentially a qualitative or a semi-quantitative test indicating the sensitivity or resistance of microorganisms to the test materials as well as the bacteriostatic or bactericidal activity of a compound. The antibacterial activity of the ligands and their chromium complexes was evaluated against Gram-positive (*B. subtilis*) and Gram-negative (*E. coli*) bacteria. The nutrient agar medium having the composition: peptone 5 g, beef extract 5 g, NaCl 5 g, agar–agar 20 g and distilled water 1000 ml was pipetted into the Petri plates. When it solidified, 5 ml of warm seeded agar was applied. The seeded agar

was prepared by cooling the molten agar to 40 °C and then 10 ml of bacterial suspension was added. The compounds were dissolved in methanol in 500 and 1000 ppm concentrations. Paper discs of Whatman No. 1 filter paper 5 mm in diameter were soaked in these solutions of varied concentrations. The discs were dried and placed on the medium previously seeded with organisms in Petri plates at a suitable distance. The Petri plates were stored in an incubator at  $28 \pm 2$  °C for 24 h. The diameters of the zone of inhibition produced by the compounds were compared with the standard antibiotic (Streptomycin). The zone of inhibition thus formed around each disc containing the test compounds was measured accurately in millimetres.

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